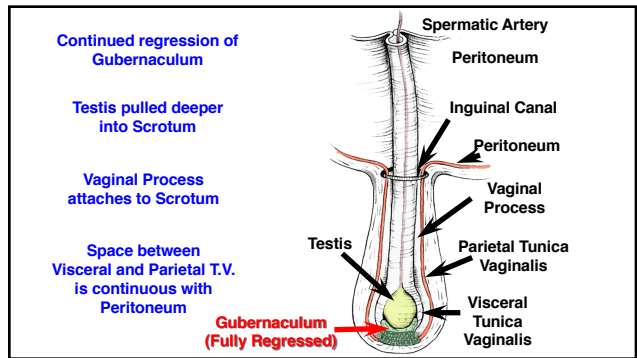


Male Reproduction

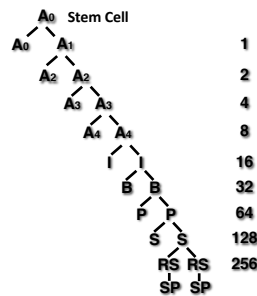
John Parrish
University of Wisconsin-Madison



Testicular Descent

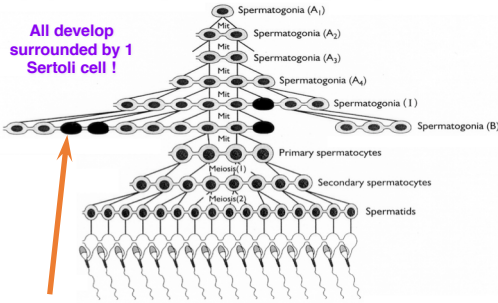
- Cryptorchid - failure of descent
- Controlling mechanisms
 - Androgen dependent
 - DHT supported
 - Estrogen inhibits
- Gene Expression
 - INSL3 - insulin like growth factor 3
 - From leydig cells
 - Great/LGR8 - receptor for INSL3
 - In gubernaculum

Potential Production of Germ Cells in Boar



França LR, Avelar GF, Almeida FFL. 2005. Theriogenology 63:300-318.

Cytoplasmic Bridges Present Among Daughter Cells



Losses During Spermatogenesis

Citation	Round Spermatids/ Sertoli Cell	% Loss
Theoretical	256	0
Franca et al., 2005	68	73.4%
Costa et al., 2013	21.5	91.6%
Parrish unpublished	30.4	88.1%

- Most losses (approx. 90%) occur during Mitotic processes
- A, I, B or Spermatogonia or Primary Spermatocyte

Boar Spermatogenesis

Transport through epididymis: 10 days

Cycle	4	RS (18.5)	ERS (17.6)	ES (16.4)	ES (16.1)	ES (15.1)	ES (14.3)	ES (12.6)	ES (11)
	3	P (28)	P (26.1)	D (24.9)	MeI/2 (24.6)	RS (23.6)	RS (22.8)	RS (21.1)	RS (19.5)
	2	PL/L (36.5)	Z (35.6)	Z (34.4)	Z (34.1)	P (33.1)	P (32.3)	P (30.6)	P (29)
	1	A (45)	A (44.1)	A (42.9)	A (42.6)	A (41.6)	B (40.8)	B (39.1)	B (37.5)
Stage	I	II	III	IV	V	VI	VII	VIII	
Days	0.9	1.2	0.3	1.0	0.8	1.7	1.6	1.0	

Duration and description of stages were from Swierstra (1968); specific cellular associations are from Frankenbuh et al. (1982) and Costa et al. (2013). 2, secondary spermatocyte; A, spermatogonium type A; B, spermatogonium type B; D, diploene primary spermatocyte; ERS, elongating round spermatid; ES, elongating spermatid; L, leptotene primary spermatocyte; Z, zygotene primary spermatocyte; P, pachytene primary spermatocyte; PL, postleptotene primary spermatocyte; RS, round spermatid; (), estimated days to ejaculation (assuming an epididymal transit time of 10 days).

Stages

- Specific cellular associations within a small segment of a seminiferous tubule
- stages are not the same length in time

Boar Spermatogenesis

Transport through epididymis: 10 days

Cycle	4	RS (18.5)	ERS (17.6)	ES (16.4)	ES (16.1)	ES (15.1)	ES (14.3)	ES (12.6)	ES (11)
	3	P (28)	P (26.1)	D (24.9)	MeI/2 (24.6)	RS (23.6)	RS (22.8)	RS (21.1)	RS (19.5)
	2	PL/L (36.5)	Z (35.6)	Z (34.4)	Z (34.1)	P (33.1)	P (32.3)	P (30.6)	P (29)
	1	A (45)	A (44.1)	A (42.9)	A (42.6)	A (41.6)	B (40.8)	B (39.1)	B (37.5)
Stage	I	II	III	IV	V	VI	VII	VIII	
Days	0.9	1.2	0.3	1.0	0.8	1.7	1.6	1.0	

Cycle

- progression through sequence of all stages
- Approx. 4 - 4.5 cycles to form spermatozoa
 > some species variation

Boar Spermatogenesis

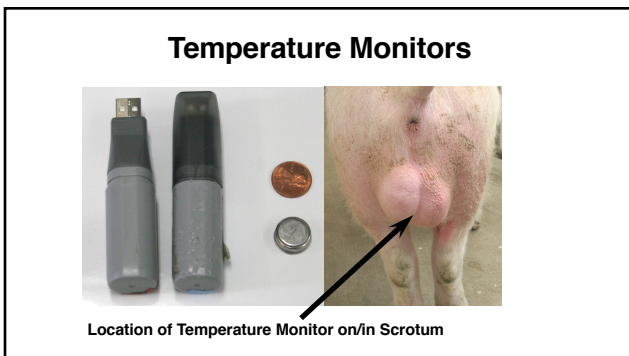
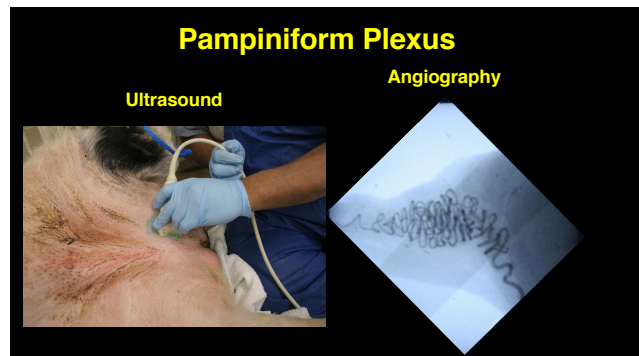
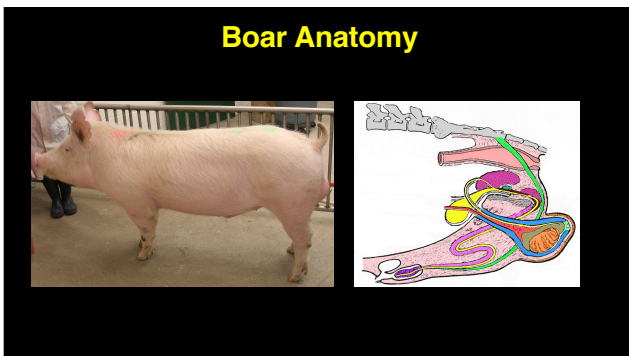
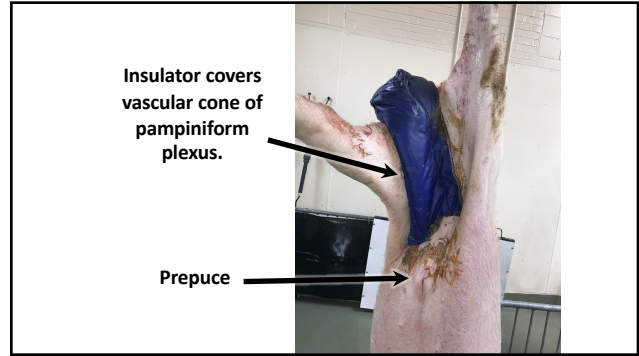
Transport through epididymis: 10 days

Cycle	4	RS (18.5)	ERS (17.6)	ES (16.4)	ES (16.1)	ES (15.1)	ES (14.3)	ES (12.6)	ES (11)
	3	P (28)	P (26.1)	D (24.9)	MeI/2 (24.6)	RS (23.6)	RS (22.8)	RS (21.1)	RS (19.5)
	2	PL/L (36.5)	Z (35.6)	Z (34.4)	Z (34.1)	P (33.1)	P (32.3)	P (30.6)	P (29)
	1	A (45)	A (44.1)	A (42.9)	A (42.6)	A (41.6)	B (40.8)	B (39.1)	B (37.5)
Stage	I	II	III	IV	V	VI	VII	VIII	
Days	0.9	1.2	0.3	1.0	0.8	1.7	1.6	1.0	

Duration of Spermatogenesis

	Bull	Ram	Boar	Stallion	Man
cycle (days)	13.5	10.4	8.5	12.2	16
Spermatogenesis	61	47	34	57	75

For Ejaculation - Add 9 - 11 days for epididymal transit



Effect of Insulation on Scrotal Temperatures

	Control ^b	Full Insulation
Surface Measurement ^a	32.1 ± 0.4°C N=5	34.0 ± 0.3°C N=5
Below Scrotal Skin ^a	34.9 ± 0.3°C N=6	38.0 ± 0.1°C N=3

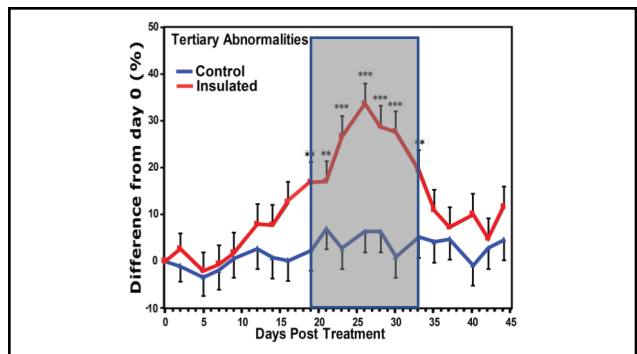
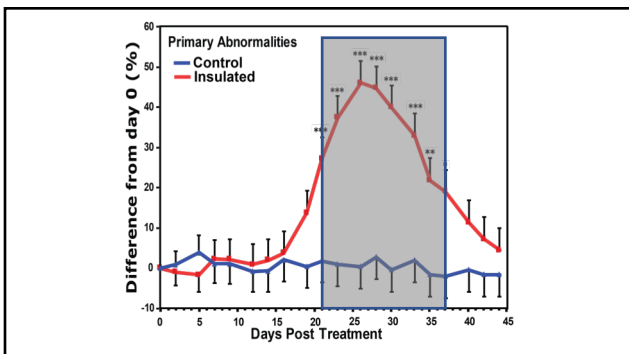
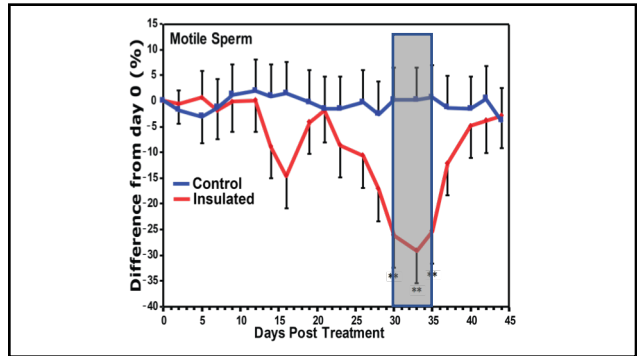
^aSurface measurement with old style USB device; Below skin with new iButton device

^bControl for surface used sham sack with no insulation; Control for below skin had not sack present.

Core body temp = 38.5°C

Experimental Procedures

- 5 control (sham), 5 insulated boars
- 48 hr insulation
- Semen collected MWF 2 weeks pre- through 6 weeks post-insulation
- Semen evaluation
 - Motility
 - Morphology
 - Sperm Nuclear Shape (Fourier Harmonic Analysis)

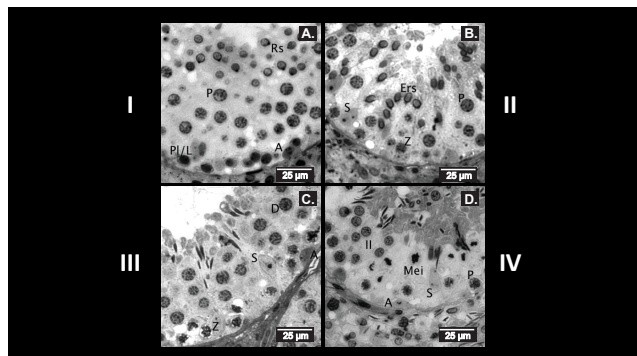


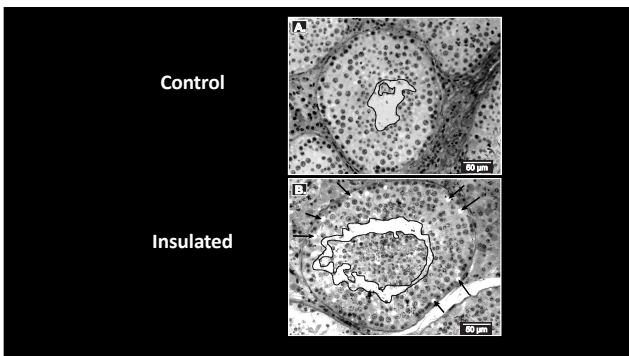
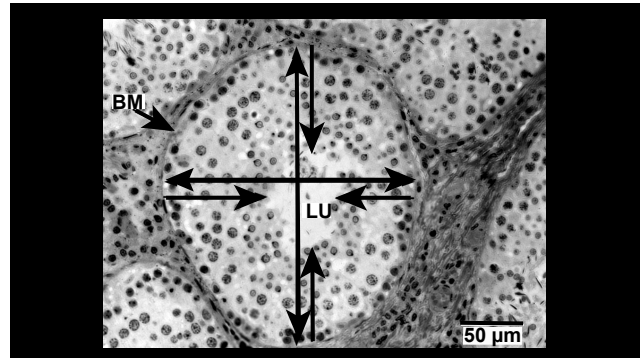
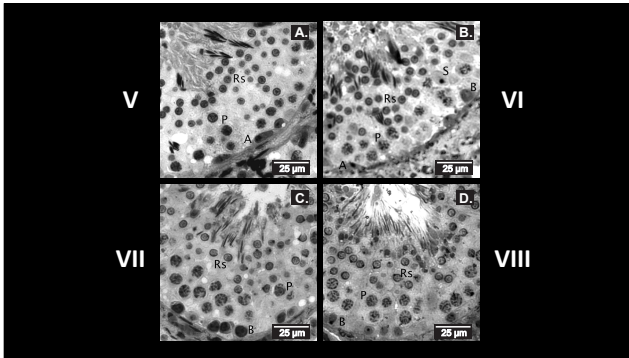
Boar Spermatogenesis

Transport through epididymis: 10 days

Cycle	4	3	2	1
Stage	RS (18.5)	ERS (17.8)	ES (16.4)	ES (15.1)
Days	0.9	1.2	0.3	1.0
Stage	ES (14.3)	RS (22.8)	RS (21.1)	RS (19.5)
Days	1.7	1.6	1.8	1.0
Stage	ES (11.4)	ES (10.8)	ES (10.2)	ES (9.6)
Days	1.7	1.6	1.8	1.0
Stage	ES (10.2)	ES (9.6)	ES (9.0)	ES (8.4)
Days	1.7	1.6	1.8	1.0
Stage	ES (9.0)	ES (8.4)	ES (7.8)	ES (7.2)
Days	1.7	1.6	1.8	1.0
Stage	ES (8.4)	ES (7.8)	ES (7.2)	ES (6.6)
Days	1.7	1.6	1.8	1.0
Stage	ES (7.2)	ES (6.6)	ES (6.0)	ES (5.4)
Days	1.7	1.6	1.8	1.0
Stage	ES (6.6)	ES (6.0)	ES (5.4)	ES (4.8)
Days	1.7	1.6	1.8	1.0
Stage	ES (5.4)	ES (4.8)	ES (4.2)	ES (3.6)
Days	1.7	1.6	1.8	1.0
Stage	ES (4.2)	ES (3.6)	ES (3.0)	ES (2.4)
Days	1.7	1.6	1.8	1.0
Stage	ES (3.6)	ES (3.0)	ES (2.4)	ES (1.8)
Days	1.7	1.6	1.8	1.0
Stage	ES (2.4)	ES (1.8)	ES (1.2)	ES (0.6)
Days	1.7	1.6	1.8	1.0
Stage	ES (1.8)	ES (1.2)	ES (0.6)	ES (0.0)
Days	1.7	1.6	1.8	1.0

Motility





The effect of scrotal insulation on scrotal and testicular parameters.^a

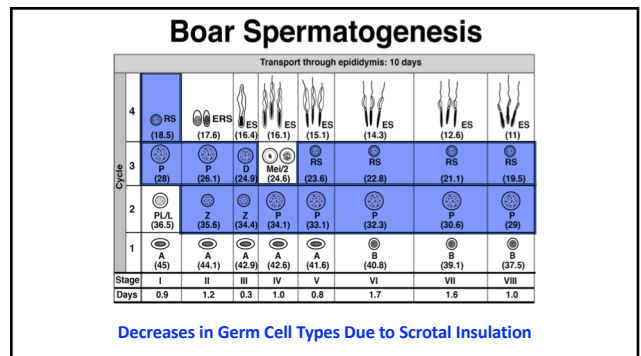
Measurement	Control	Insulated	Pooled SE
Scrotal temperature (°C)	31.1	33.9*	0.6
Height of seminal epithelium (µm)	64.4	55.1*	1.22
Amount of debris in the lumen (%)	26.0	45.5*	2.8

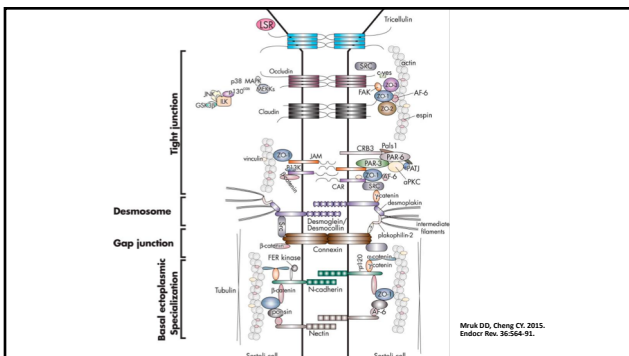
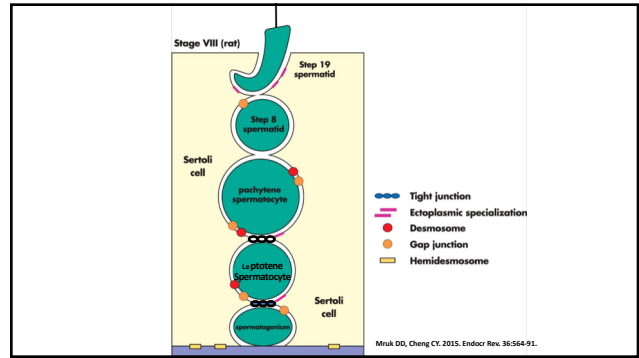
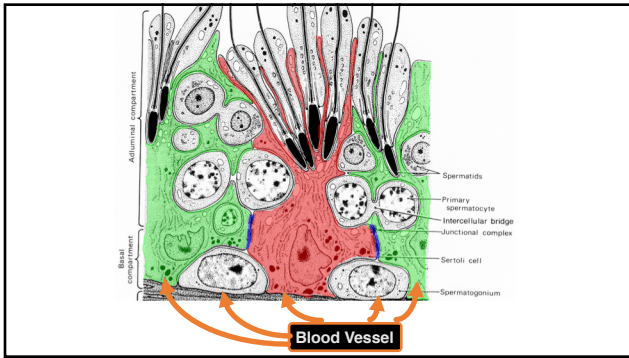
^aData are the mean ± sem for 5 control (sham insulated) and 5 insulated boars.
* Indicates a difference within the row, p < 0.05.

The effect of scrotal insulation on cells within the seminiferous tubules.^a

Cell Type	Control	Insulated	Pooled SE
Sertoli cells	0.9	1.0 nd	0.1
A spermatogonia	2.2	1.9 nd	0.1
B spermatogonia	5.7	5.7 nd	0.4
Prelep./lep. 1° spermatocytes	11.2	9.0	0.8
Zygotene 1° spermatocytes	13.2	10.7 *	0.5
Pachytene 1° spermatocytes	13.6	10.4 *	0.4
Diplotene 1° spermatocytes	14.2	10.6 *	0.9
Meiotic bodies	5.2	4.8 nd	0.7
2° spermatocytes	14.3	10.3 nd	1.7
Round spermatids	33.0	28.0 *	1.4
Elongating round spermatids	29.1	22.3 nd	2.7

^aData are the mean ± sem for 5 control (sham insulated) and 5 insulated boars.
* Indicates a difference within the row, p < 0.05.
ndIndicates there was no difference within the row, p > 0.05.





Mechanisms of Heat Stress Damage to Spermatogenesis

- **Dysregulation of BTB junctional complexes** (Cai et al., 2011)
 - Observed in Mouse
 - Normal sequence
 - Breakdown of tight junctions and ectoplasmic specializations
 - Reassembly basal to primary spermatocyte with recycled proteins
 - Suggest relocating BTB components disrupted
 - BTB must recover as sperm are produced
 - Mice – BTB is damaged but recovers and involves Androgen signaling
 - Data in boar lacking

Mechanisms of Heat Stress Damage to Spermatogenesis

- **Apoptosis** (Durairajanayagam et al., 2015)
 - Rats – germ cells undergo apoptosis in response to heat stress
 - Consistent with gene expression and proteins in sperm
 - DNA repair occurs following DNA damage of apoptosis
 - Only partial however
 - **Stress Pathways** (Durairajanayagam et al., 2015)
 - Increased lipid peroxidation, reduced superoxide dismutase and catalase activity.
 - Increased ROS
 - **DNA repair reduced** (Tramontano et al., 2000)
 - **Intrinsic apoptosis – Mitochondria**
 - **Extrinsic apoptosis – Fas/FAS-ligand signalling**

Mechanisms of Heat Stress Damage to Spermatogenesis

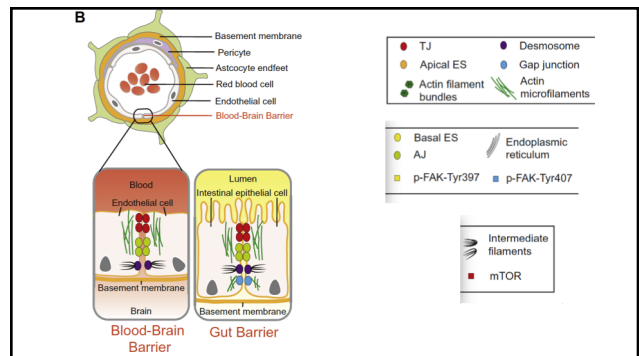
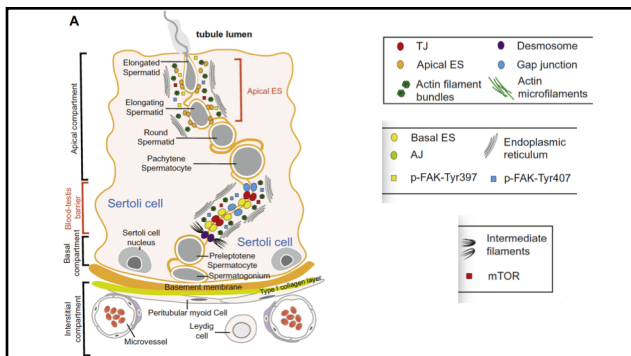
- **Autophagy** (Durairajanayagam et al., 2015)
 - Flag cells for ubiquitin conjugation
- **Cytoplasmic droplet retention** (Cooper, 2011)
 - Involved in sperm volume regulation upon ejaculation
 - Retained droplet can not function and sperm compromised

Preventing Heat Stress Damage

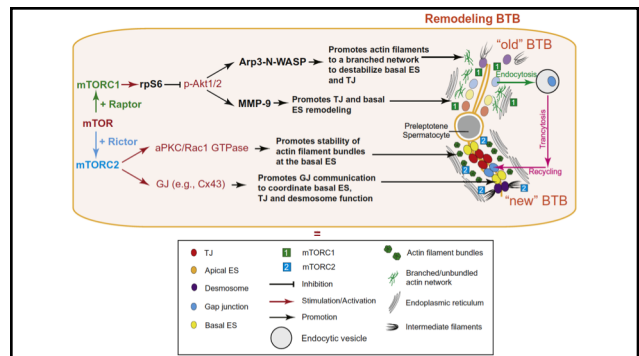
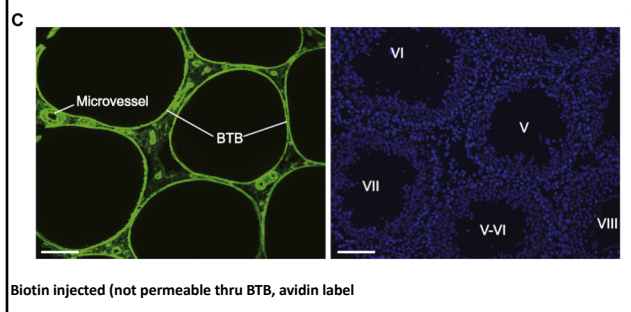
- **Cooling**
 - Cool-cell technology – reduces air temp. only 10°F
 - Air conditioning – cost expensive
 - Scrotal mists/drips – requires constant air flow
 - Humans – loose cloths, air conditioning
- **Naturally resistant male**
 - Livestock - How to identify?
- **Impact BTB**
 - Androgens, hCG, GnRH analogues
 - FSH, eCG
- **Inhibit ROS and DNA damage**
 - Vitamin E, catalase, antioxidants all given to boar

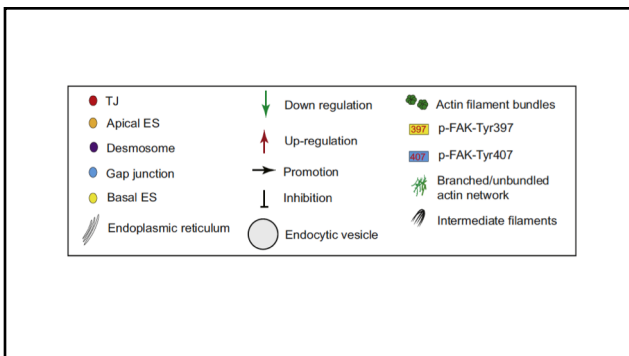
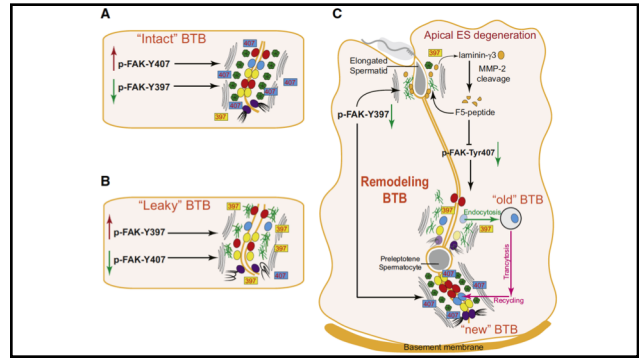
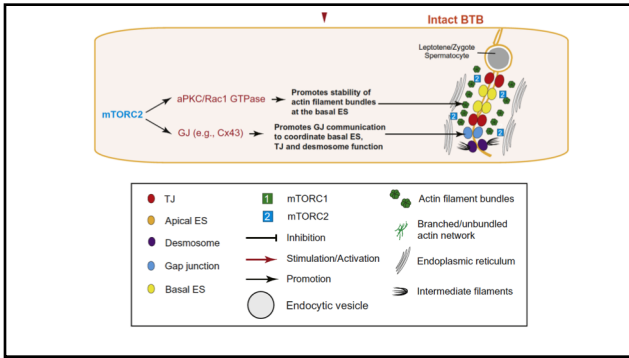
The Blood Testis Barrier

- What is it?
- What is its role?
- What are other barriers in the body similar to the BTB?
- As pre-leptotene primary spermatocyte moves across BTB, to form Leptotene primary spermatocyte what happens?
- What is the role of mTORC1 and TORC2?
- What is the role of p-FAK-Y407 and p-FAK-Y397?



Does not become leaky as pre-leptotene to Leptotene movement.





The Blood Testis Barrier and apical specializations are complex !!!